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Intranasal Dexamethasone: a New Clinical Trial For The Control of Inflammation and Neuroinflammation in Covid-19 Patients

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1 INTRANASAL DEXAMETHASONE: A NEW CLINICAL TRIAL FOR THE CONTROL
2 OF INFLAMMATION AND NEUROINFLAMMATION IN COVID-19 PATIENTS

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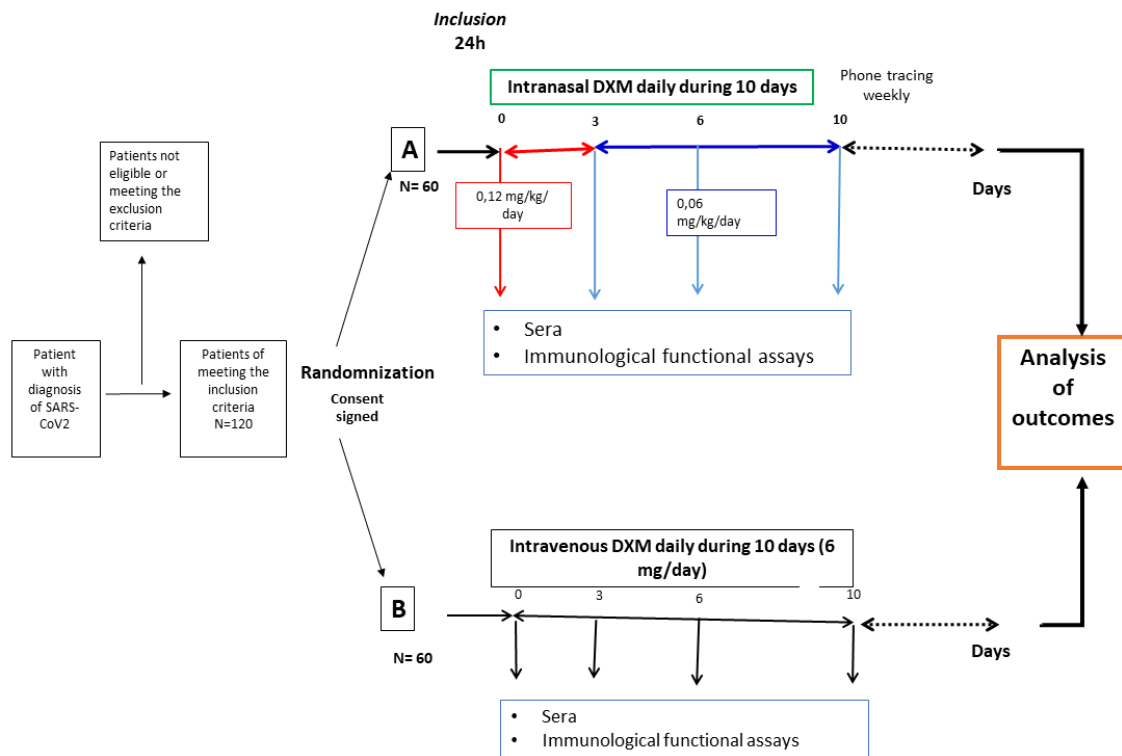
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57 Graphical Abstract



58 Highlights

- 59 • REVIVAL is a controlled, open-label multicentric study to compare the standard low
60 doses of intravenous dexamethasone with low doses weight-adjusted of intranasal
61 dexamethasone
- 62 • Intranasal dexamethasone can reach more effectively than intravenous the respiratory
63 tract
- 64 • Intranasal dexamethasone can reach the central nervous system in therapeutic
65 concentrations even at low doses
- 66 • REVIVAL aims to add to the control of systemic inflammation, the control of
67 neuroinflammation to reduce central failures and sequelae

68 **Abstract**

69 COVID-19 has produced more than 176 million infected individuals and almost 3.2 million
70 deaths worldwide. The infection results in a dysregulated systemic inflammation, multi-
71 organ dysfunction, and critical illness. Cells of the central nervous system (CNS) are also
72 affected triggering a dysregulated neuroinflammatory response.

73 Low doses of glucocorticoids (GCs) orally or intravenously administered has been proved to
74 reduce mortality of moderate and severe COVID-19 patients. However, low doses
75 administered by those routes do not reach therapeutic levels in the CNS. In contrast, if
76 dexamethasone is administered by the intranasal route can result in therapeutic doses in the
77 CNS even at low doses of the GC.

78 **Methods:** This is an approved multicentric randomized controlled protocol to compare the
79 effectiveness of low doses of intranasal dexamethasone versus intravenous administered in
80 adult moderate and severe COVID-19 patients. The protocol is conducted in five health
81 institutions in Mexico City. A total of 120 patients will be randomized in two groups
82 (intravenous vs intranasal) at 1:1 ratio, both groups will be treated with these dexamethasone
83 schemes for 10 days. The primary outcome of the study will be clinical improvement, defined
84 as a statistically significant higher reduction in the NEWS-2 score in intranasally versus
85 intravenously dexamethasone treated patients. The second outcome will be the reduction in
86 mortality during hospitalization.

87 **Conclusions:** This protocol is currently undertaken to improve the efficacy of the standard
88 therapeutic dexamethasone regimen for-moderate and severe COVID-19 patients.

89 **Trial registration:** ClinicalTrials.gov identifier: NCT04513184 Registered November 12,
90 2020 and was approved by COFEPRIS with identifier DI/20/407/04/36. People are currently
91 being recruited.

92 **Keywords:** Dexamethasone, intranasal administration, inflammation, neuroinflammation,
93 COVID-19

94

95 **Background**

96 So far, the outbreak of COVID-19 has caused more than 176 million infected individuals and
97 almost 3.2 million deaths worldwide (<https://coronavirus.jhu.edu/map.html>) with a current
98 global case-fatality ratio of 2.1%, the most affected geographic region are the Americas with
99 a case-fatality ratio of 2.6%.

100 Several factors predict a poor outcome for COVID-19 patients, such as comorbidities
101 (diabetes, hypertension, obesity) and aging with an underlying dysregulated inflammatory
102 response¹. Other relevant factors include SARS-CoV-2 neurotropism/neuroinvasiveness²⁻⁹. In
103 fact, the viral RNA was observed in the brain of patients that deceased by severe acute
104 respiratory syndrome due to COVID-19 infection¹⁰⁻¹². Likewise, it was reported evidence
105 of astrocytic activation and neuronal damage in severe COVID-19 patients, which present
106 elevated plasmatic levels of GFAP and NfL¹³. Other authors have evaluated astrocytes¹⁴
107 and neurons in 2D or 3D cultures showing an extensive infection^{15,16}. The infection of cells
108 of the Central Nervous System results in the expression of PAMPs and DAMPs that trigger
109 a neuroinflammatory response. The exacerbated systemic inflammation with the
110 consequent breakdown of the blood-brain barrier and the migration of cells and peripheral
111 inflammatory mediators also contribute to increase to the in situ generated
112 neuroinflammatory response. Together, this dysregulated and sustained neuroinflammation
113 can add to peripheral damage, central (CNS) damage, which may contribute to the multi-
114 organ dysfunction and death^{10,12}.

115

116

117 **Natural history of SARS-CoV-2 infection**

118 A clinical staging system has been proposed in SARS-CoV-2 infection as follow, early
119 infection (Stage I, mild), pulmonary involvement (Stage IIa, moderate) without hypoxia, or
120 with hypoxia (Stage IIb), and finally Stage III (systemic hyperinflammation) ¹⁷ (Figure 1).

121 After exposure to SARS-CoV-2, virus gains host access through the nasal cavity and
122 respiratory airway. During early infection (Stage I), mild and non – specific symptoms may
123 be observed (fever, malaise, and asthenia), upon this prodromic phase virus binds its target
124 ACE2, TMPRSS2 ^{18, 19} and more recently NRP-1 ^{20,21}. These receptors are highly present on
125 several tissues including the olfactory neuroepithelium (less in the sensitive olfactory
126 neurons) and lung ¹⁹⁻²², consequently, the infection can be established in the lungs (Stage II)
127 and lead to viral pneumonia, cough, and fever with or without hypoxia. Here the SARS-CoV-
128 2 PAMPs will be recognized by TLR3, TLR7 and TLR8 in the endosome but also in the RIG-
129 1 like receptor in the cytosol ²³. The virus can also reach the CNS through the olfactory and
130 trigeminal nerves terminal. Once in the CNS it can infect and damage endothelial, pericytes
131 and neural cells that expressed ACE2, NRP-1 receptors ^{20, 21} promoting neuroinflammation
132 (Figure 1). CNS viral involvement is related to headache, dizziness, and ataxia, but infection
133 also may progress to the whole brain including the brainstem ^{5, 6}. Finally, in a minority of
134 infected-patients disease progresses to Stage III where a hyperinflammatory syndrome (the
135 sustained production of proinflammatory cytokines including IL-1 β and TNF α) is observed,
136 with mitochondrial and lysosomal damage, expressing elevated proinflammatory cytokines,
137 reactive oxygen species (ROS), and the hyperactivation of P2X7 receptors. These processes
138 induce inflammasome activation (which increased IL-6 levels) and lead to pyroptosis which

139 determines a persistent inflammatory cycle by disseminating viral antigens and RNA in the
140 circulation. Thereafter, it is possible the generation of immune complex and its deposition in
141 target organs ²³⁻²⁵. During this phase, sustained neuroinflammation may exacerbate the
142 neuronal injury, therefore spreading damage and contributing towards central respiratory
143 failure besides other signs of systemic organ involvement resulting in multi-organ
144 dysfunction ¹⁷.

145 A crucial strategy to treat COVID-19 patients seems to be the control of neuroinflammation
146 and systemic inflammation. For this purpose, it is important to consider how the virus invades
147 the human organism. The most frequent form is the intranasal route which allows a direct
148 access to both, the respiratory and the central nervous systems through neural pathways ^{5; 15-}
149 ¹⁸. Coronaviruses including SARS-CoV-2 can infect brainstem neurons associated with
150 cardio-respiratory control, which induces central alterations of pulmonary function ^{5; 26-29}. In
151 fact, COVID-19 neurological clinical symptoms particularly nausea, vomiting, and dysgeusia
152 appear to involve the dorsal vagal complex (DVC) and the nucleus tractus solitary (NTS)
153 linked to the control of several autonomic functions ²⁶. The NTS is a well-known target of
154 neuro-immune activation ³³, and its ascending projections reach the hypothalamus
155 (hypothalamic paraventricular nucleus) involved in the HPA axis activation while other NTS
156 projections come to the rostral ventrolateral medulla (RVM), which controls respiratory and
157 cardiovascular functions ³⁴.

158 The viral infection in respiratory and central nervous system cells promotes the expression
159 of pathogen-associated molecular patterns (PAMPs) and damage-associated molecular
160 patterns (DAMPs) signals that in turn trigger inflammasome and oxidative stress ^{23,35}. Later

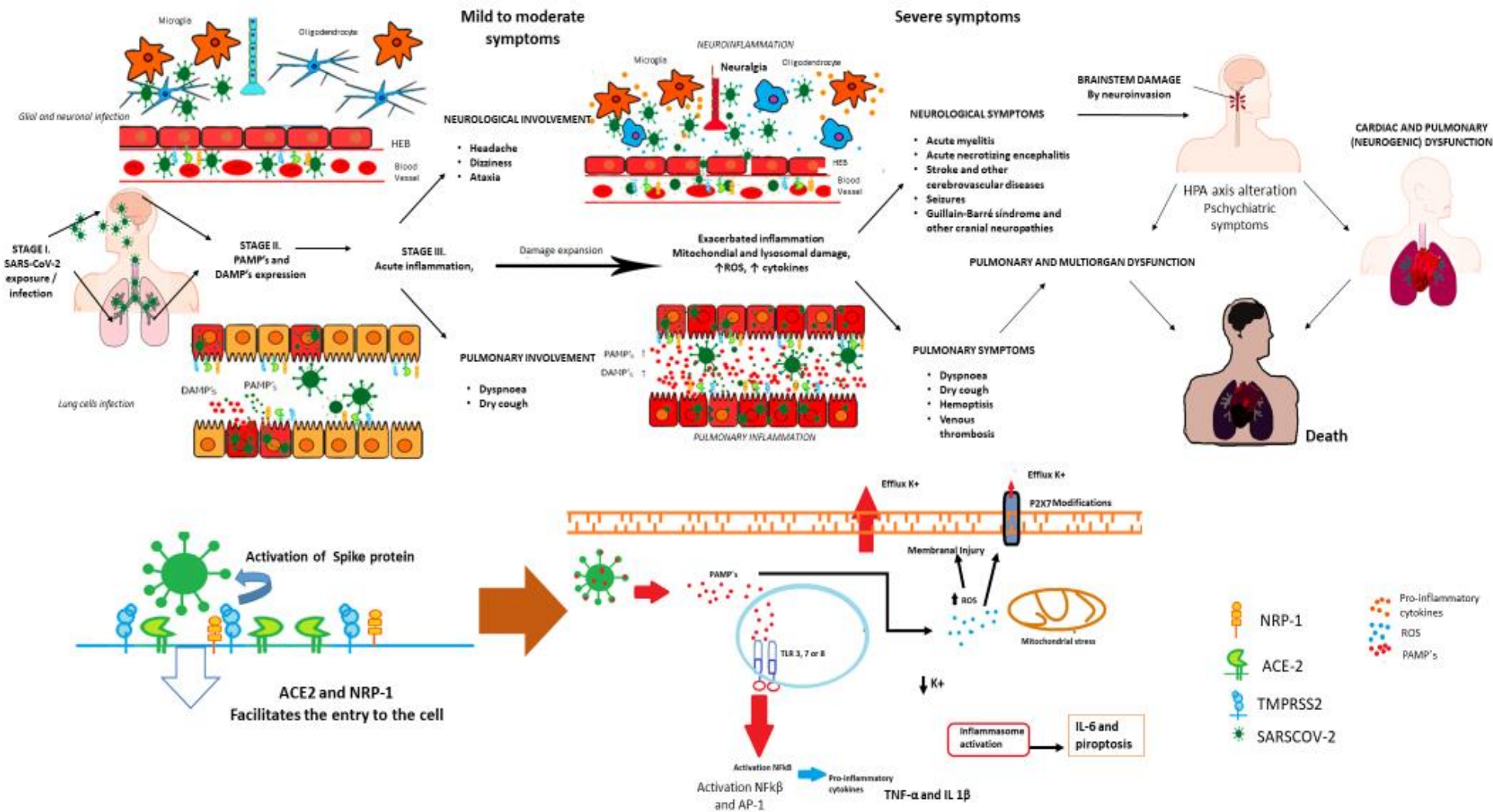
161 during infection, inflammatory response may become dysregulated extending the initial
162 damage caused by the infection.

163 **Adrenal affection in SARS-CoV-2 infection**

164 Critically ill-patients of different pathologies frequently show adrenal insufficiency which
165 may increase morbidity and mortality ^{36, 37}. COVID-19 might affect the hypothalamic-
166 pituitary-adrenal (HPA) axis as well. Hypothalamic and hypophysis tissues do express ACE2
167 and can therefore be viral targets ³⁸. The virus may directly damage the hypothalamus as well
168 as the pituitary leading to hypothalamo-pituitary dysfunctions. In fact, since SARS outbreak
169 in 2003, it was observed that coronavirus affects the HPA axis, and vasculitis was
170 demonstrated by autopsy studies in several organs including adrenal glands, particularly
171 adrenal cortical cells undergo degeneration and necrosis ³⁹. Although the full spectrum of
172 COVID-19 endocrinological manifestations within long-term is still unclear, several
173 endocrine alterations have been reported in SARS survivors, as well as hypocortisolism, and
174 hypothyroidism, and low levels of dehydroepiandrosterone, which suggested a transient
175 hypothalamic-pituitary dysfunction ⁴⁰. Recently, an Arabian study in 28-patients reported the
176 adrenal response to an acute COVID-19 infection, the median morning cortisol level was 196
177 (31-587) nmol/L, the ACTH median level of 18.5 (4-38ng/L). Interestingly, severe forms
178 patients had lower cortisol and ACTH ⁴¹. In addition, in other autopsy studies, edema,
179 neuronal degeneration and evidence of viral genome were found in the hypothalamus ⁴² Thus,
180 in the presence of subacute thyroiditis or adrenal insufficiency, corticosteroid therapy should
181 help in reduced high amounts of thyroid hormones, and replace adrenal function, improving
182 the evolution of these patients, regardless the route of administration.

183 **Rationale**

184 Dexamethasone sodium phosphate (ALIN, injectable solution. Chinoin Laboratory) is a
185 highly soluble glucocorticoid with a neutral pH 7-8.5, which did not injury the nasal mucosa.
186 This synthetic steroid is an anti-inflammatory and immunomodulator drug that inhibits
187 prostaglandins and leukotrienes synthesis, platelet activation, and coagulation through
188 regulation of transcriptional factors such as NFK- β y AP-1 ^{43,44}. In addition, it can sensitize
189 the cells to extracellular ATP during NLRP3 induction, which enhances the release of
190 proinflammatory molecules ⁴⁵. In addition, it has been reported that DXM exerts important
191 neuroprotective effects as rescue the neurovascular integrity during neuroinflammation ⁴⁶.



193 Figure 1.

194 **Dexamethasone a potent anti-inflammatory drug**

195 Considering that complications of COVID-19 result from exacerbated and uncontrolled
196 peripheral inflammation and neuroinflammation, derived from the so-called cytokine storms,
197 at least three important and key points have been considered in the use of DXM for the
198 treatment of victims of the Coronavirus: the timing, the dose, and the route of administration
199 of the steroid. First, the drug would not be applied from the beginning of the infection, the
200 time at which the inflammation favors the host. It should be given to promote the installation
201 of an adaptive immune response and thus control the infection. A low dose of DXM (6 mg
202 per patient for 10 days) applied to quickly and effectively control pulmonary inflammation
203 with minimal negative side effects ⁴⁷. In addition, the intranasal route would allow direct
204 access of the DXM to the CNS, thereby controlling the sustained neuroinflammation
205 provoked by damage to infected astrocytes, neurons and microglia during the progression of
206 COVID that cause the fatal central respiratory and cardiac failure in these patients.

207 It is well known that drugs administered intranasal usually permit higher bioavailability in
208 CNS without the need of BBB pass or hepatic degradation, in comparison with similar
209 intravenous doses administered in experimental models ^{55, 56-58}. In addition, the
210 administration of DXM by this route induces an inflammatory control by arriving directly to
211 the respiratory system, more effectively and quickly than by using intravenous route ⁵⁶⁻⁵⁹.
212 DXM prevents the binding of ACE2 to spike protein of SARS-CoV-2 and can bind LYS353,
213 an active residue of RBD ⁶⁰, and reduces ACE2 expression in several types of cells by
214 suppressing type I interferon expression ⁶¹, can also downregulate neutrophils extracellular
215 traps, possibly through Toll-like receptor regulation ⁶². It is known that hyper inflammation

216 is related to high levels of NETs which is related to ARDS in which neutrophilia predicts
217 thrombosis and poorer outcomes ^{63, 64}.

218 **METHODS**

219 **Trial design**

220 The “REVIVAL trial” an interventional study, phase 2, multicentric randomized controlled
221 in adult patients with confirmed COVID-19 diagnosis was designed to evaluate the efficacy
222 of low doses of intranasal DXM compared to intravenous administration in patients of five
223 COVID-19 referral centers in Mexico City. This protocol is supported in part by the
224 Institutional grant "Programa de Investigación para el Desarrollo y la Optimización de
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227 grant provided by the Mexican Ministry of Foreign Affairs (Secretaria de Relaciones
228 Exteriores) and Mexican Agency for International Development Cooperation (AMEXCID)
229 with identifier: 318.01 fund MEX-CHI. This trial is being coordinated at the Department of
230 Immunology of the Biomedical Research Institute, UNAM.

231 **Settings**

232 This clinical trial is being conducted at the following Institutions “Hospital General de
233 México Dr. Eduardo Liceaga”, “Instituto Nacional de Neurología y Neurocirugía Manuel
234 Velasco Suárez”, “Instituto Nacional de Cardiología Ignacio Chavez”, “COVID-19 unit at
235 Citibanamex” and “Hospital Central Militar” all of them in Mexico City.

236

237 **Eligibility criteria**

238 Inclusion criteria includes patients of both sexes, (non-pregnant female) 18 years of age and
239 under 90 years, with presumptive SARS-CoV-2 infection with more than 5 days of clinical
240 evolution and with moderate to severe symptoms requiring oxygen support or high flux
241 mechanical ventilation (NEWS-2 \geq 5), abnormal CT- chest scan CO-RADS >3 . Patients
242 diagnosed with atypical pneumonia, confirmed by chest images and oxygen saturation
243 (SpO₂) less than 93% in ambient air or when a ratio of the partial pressure of oxygen (PaO₂)
244 and the fraction of inspired oxygen (FiO₂) (PaO₂: FiO₂) was 300 mm Hg or less, and a
245 confirmatory RT- PCR SARS-CoV-2 positive test. These patients will be allocated into the
246 experimental group or the control group in a ratio 1:1 (two arms) (Fig. 2) according to the
247 randomization.

248 Exclusion criteria includes patients with RT-PCR SARS-CoV-2 negative test, those
249 receiving previously GCs at high doses, by oral or intravenous administration, or severely
250 immunosuppressed as in AIDS, pregnancy; autoimmune disease patients as well as those
251 who have received outpatient treatment with steroids for more than 72 hours prior to hospital
252 admission, older than 90 years, or with DXM allergy, risk for glaucoma or recurrent
253 respiratory diseases.

254 Elimination criteria will be considered in case of voluntary withdrawing or lacking informed
255 consent, or imminent risk of death within 48 hrs.

256 The pharmacovigilance staff of each hospital will perform a continuous monitoring each 72
257 hours during the period of study (including all adverse events).

258

259 **Interventions**

260 **Groups and comparators**

261 The study will be carried out in two groups, group A (experimental) that will receive
262 intranasal DXM, and group B (Control) that will receive intravenous DXM (Fig.2), based on
263 the previously reported data, where the intranasal administration can reach the brain and
264 bloodstream more quickly and efficiently⁵⁶⁻⁵⁹. Group A will receive daily intranasally DXM
265 at a dose of 0.12 mg / kg for the first three days, that will be followed by seven days at a dose
266 of 0.06mg / kg. Group B will receive daily 6mg intravenous DXM. In both groups, a close
267 follow-up will be done by the pharmacovigilance staff every 72 hours, they will assess
268 whether it is appropriate for the patients to continue within the protocol.

269 **Procedures**

270 A double follow-up form (written and online) will be filled for each patient, and completed
271 at the end of treatment or fatal outcome after randomization, whatever occurs first. Besides
272 a daily clinical evaluation, blood and saliva samples will be collected every third day during
273 the whole treatment period, to perform ancillary tests as SARS-CoV-2 viral load, functional
274 immunological assessment (lymphocyte cytometry, cytokines / chemokines profile), as well
275 as cortisol levels, among other analysis. All human samples will be stored at -70°C until use.
276 All patient's personal data and medical information will be treated in a strictly confidential
277 way. Only the lead investigator and the hospital coordinator investigators will have access.

278

279

280 **Participants**

281 The sample includes 120 adult patients between 18 and 90 year-olds, both sexes with
282 moderate and severe forms of COVID-19.

283 **Sample size and Randomization**

284 The sample size was calculated with EPIDAT version 3.1.2 software, with the option
285 “Sample size and surveillance curves” with an estimation of 50% increase in the proportion
286 of patients free of mechanical ventilation [intranasal DXM 70% vs intravenous DXM 45%].
287 This value was estimated based on the data of the COVID-19 patients registered in Mexican
288 hospitals with a confidence of 95%, power of 80% and proportion of losses of 10%, with
289 these characteristics is obtained 60 per group. The randomization will be making with Sealed
290 Envelope software. This software is a freeware [Online] available from:
291 <https://www.sealedenvelope.com/simple-randomiser/v1/lists> [Accessed 5 May 2020]. This
292 study is a multicenter randomized controlled trial. (Fig. 2)

293 **Confidentially**

294 Each patient who agrees to participate in the protocol will be assigned an identification
295 number, which will be used to check out throughout the procedure. This code identifies the
296 hospital of origin and the patient identification number. All the information collected during
297 the procedure will be confidential and used for the research purpose only and follow up for
298 any adverse effect.

299

300

301 **Outcomes**

302 The expected primary outcome is clinical improvement, defined as a two-point improvement
303 of ordinal scale regarding the initial NEWS-2 score. The secondary expected outcome
304 includes a reduction in mortality that will be follow-up during treatment (after
305 randomization), as well as a reduction of the time required for mechanical ventilation and the
306 length time of patient's stay in hospital. Viral load, several other physiological parameters
307 and the immune-inflammatory profile will also be evaluated before and after treatment (see
308 above).

309 **Data collection and management**

310 The patient receives an informed consent letter, where the characteristics of the procedure
311 are detailed, if he accepts, the letter will be signed and the patient will be randomized; saliva
312 and nasopharyngeal sample will be taken to know the viral load and treatment will begin as
313 indicated in figure 2; a clinical history will be made based on the initial results and physical
314 inspection of the patient.

315 The samples taken will be sent for specialized analysis following standardized operating
316 procedures (SOP's) for the analysis.

317 **Plans to promote participant retention and complete follow-up**

318 All participants in the research protocol will receive specialized medical care, by monitoring
319 continuously clinical, neurological, and neuropsychological studies. These evaluations will
320 be carried out to monitor the evolution of the disease at 1, 3, 6, and 12 months after COVID-
321 19. Those participants presenting some functional decline post COVID will be received

322 medical treatment and neurorehabilitation.

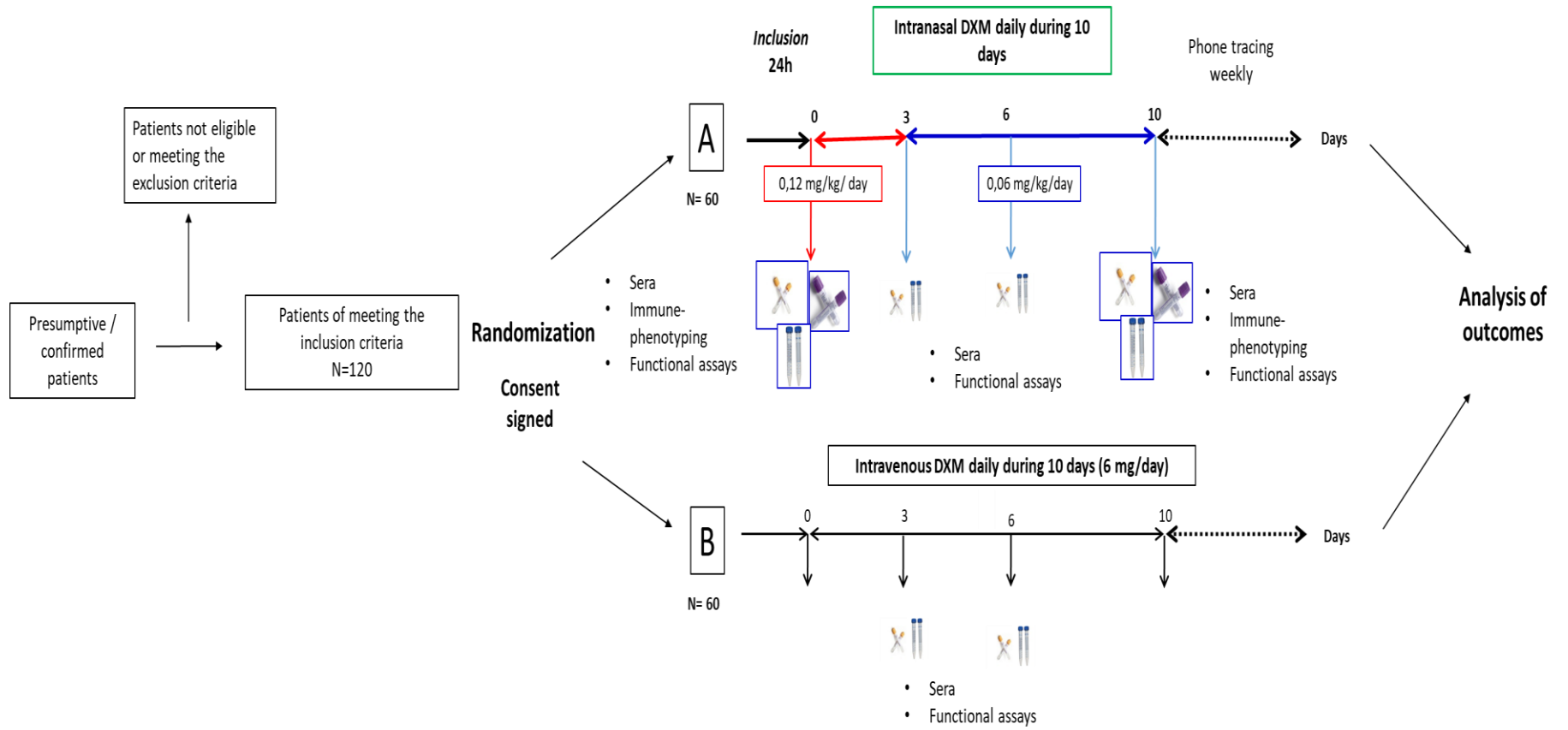
323 Likewise, for patients who present an adverse effect or health problem during its participation
324 in the dexamethasone treatment study or derivate to it upon hospitalization, the General
325 Hospital of Mexico Dr. Eduardo Liceaga will take care of the necessary treatment and/or care
326 until their resolution. In addition, patients will be monitored every 3 months for 1 year after
327 the study

328 **Management**

329 The information collected during the procedure will be documented physically and digitally
330 in an exact and precise way. Each complete patient report will be used by researchers in
331 conjunction with the molecular and immunological tests to analyze the outcomes. The
332 information collected will be treated as confidential, and only the global results will be
333 published without showing the names of the patients, in case the data is required, the
334 information can be request to the researchers with valid reasons.

335 **Analysis of outcomes**

336 A database will be built, and a descriptive statistic will be performed. The data distribution
337 will be analyzed and compared with DXM route of administration with a multivariable
338 analysis: nested ANOVA with repeated measures and Markov test. The analysis will be done
339 with a R software (4.0.0, Arbor Day). A statistical difference with $P < 0.05$ will be considered
340 significant.



341

342 **Figure 2.**

343

344 **5. Conclusions**

345 Intranasal DXM at low doses could be a more effective therapeutic option to control
346 inflammation and neuroinflammation during ARDS in severe and critical forms of SARS-
347 CoV-2 infection. In addition, it could aid the HPA axis upon this severe stress condition.
348 DXM in low doses applied by systemic route although beneficial for COVID-19 patients,
349 cannot reach effective therapeutic concentration in the CNS to control neuroinflammation.
350 In contrast, intranasal administration of DXM is highly effective to control
351 neuroinflammation as demonstrated in experimental models of several inflammatory
352 conditions⁴⁴⁻⁴⁷. Therefore, in the REVIVAL trial clinical protocol, we propose boosting the
353 effect of DXM treatment at low doses in COVID-19 through an intranasal route of
354 administration to reach CNS at therapeutic doses that may effectively reduce the morbidity
355 and mortality in severe or critical COVID-19 patients, even more than that reported data in
356 the RECOVERY trial.

357 A randomized study in hospitalized COVID-19 patients (moderate and severe forms), the
358 intranasal DXM at low doses (clinicaltrials.gov id: NCT04513184) is being tested. The
359 clinical evolution and respiratory parameters of the patients receiving intranasal DXM
360 (experimental treatment) is compared with recommended treatment of 6 mg of intravenously
361 DXM (<https://www.covid19treatmentguidelines.nih.gov/>). Considering the prevalence of
362 metabolic syndrome and obesity in Mexico, a therapeutic scheme weight-adjusted at low
363 dose is being applied i.e., three-day schedule of 0.12 mg/kg and 7 days at 0.06 mg/kg. If the
364 current approach results less prone to adverse effects but enough to reach CNS and control
365 neuroinflammation as we hypothesized, there will be direct interest to extent this protocol to

366 several COVID hospitals of the National Healthy System in Mexico. In addition, it will be
367 mandatory to increase the initial sample size (preliminary results) to publish it and share it
368 with the International scientific community.

369 **6. Declarations**

370 **6.1 Study Status**

371 The study was registered under the platform Clinical Trials from NCBI in August 2020, and
372 was approved by COFEPRIS in Mexico with identifier DI/20/407/04/36. People are currently
373 being recruited.

374 **6.2 Ethics declarations**

375 This study was reviewed and approved by the Committees of Ethics, Research and
376 Biosecurity of the five Hospitals committees. Hospital General de México “Dr. Eduardo
377 Liceaga” (DI/20/407/04/36), Instituto Nacional de Neurología y Neurocirugía (INNN 31/20)
378 and Instituto Nacional de Cardiología (INCICH: 20-1167), temporary unit for COVID
379 Citibanamex, and Hospital Central Militar (FM/DI/107/SR/2020), and is approved for
380 COFEPRIS with identifier DI/20/407/04/36.

381 All participants will provide a written informed consent before enrollment and all the work
382 will be conducted according to the Helsinki statements.

383 **6.3 Availability of data and materials**

384 Data and materials are not available at this moment, because the work being considered is
385 the first approach to a clinical trial currently started. When the study will be completed, the

386 dataset obtained and analyzed will be available from the corresponding author only by
387 reasonable request.

388 **6.4 Funding**

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394 Agency for International Development Cooperation (AMEXCID) with identifier: 318.01
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396 **6.5 Conflict of interest**

397 The authors have no other relevant affiliations or financial involvement with any organization
398 or entity with a financial interest in or financial conflict with the subject matter or materials
399 discussed in the manuscript apart from those disclosed.

400 **7. Author's contributions**

401 Study concept and design: GC, ES, HB, MR, JH, MCC

402 Data adquisition and interpretation: GC, ES, HB, MR, JH, MCC, AJR, DAM, MFMM, LVTA,
403 RLBC, RMW, LERG, KIC, EGV, MRC, YL, MLHM, MLH, KMQ, ASM, SHD, IGRZM, AMC,
404 INMS, EBS, AFP, MJFM, PSHH, JC, LH, NAF, MH, MPT, GM, HJ, EEA, GR, ROA, SOF, SRM,
405 JAHA, JCT, AFR, HB, MCR, RJB, GS, JLA, GF, JPL.RIAR, DMR, LRRRA, RAAB

406 Manuscript drafting: GC, ES, HB, MR, JH, JAHA, MCR, RJB, GS, JLA, GF, JPL

407 Critical revision of the manuscript for important intellectual content: GC, ES, HB, MR, JH, JAHA,
408 MCR, RJB, GS, JLA, GF, JPL

409 **8. Consent for publication**

410 Not applicable

411 **9. Acknowledgements**

412 Not applicable

413 **10. List of abbreviations**

414 **DVC:** Dorsal Vagal Complex

415 **NTS:** Nucleus Tractus Solitary

416 **HPA:** Hypothalamic Pituitary Adrenal axis

417 **RVM:** Rostral Ventrolateral Medulla

418 **DAMP:** Damage-Associated Molecular Patterns

419 **PAMP:** Pathogen-Associated Molecular Patterns

420 **DXM:** Dexamethasone

421 **GCs:** Glucocorticoids

422 **ACE2:** Angiotensin-Converting Enzyme 2

423 **TMPRSS2:** Transmembrane Protease Serine 2

424 **CNS:** Central Nervous System

425 **ROS:** Reactive Oxygen Species

- 426 **NFK-B:** Nuclear Factor K beta
- 427 **AP-1:** Activator Protein 1
- 428 **ARDS:** Severe Acute Respiratory Distress Syndrome
- 429 **BBB:** Blood Brain Barrier
- 430 **NET:** Neutrophil Extracellular Traps
- 431

432 **Figure Legends**

433 **Figure 1.** Inflammatory phenomenon associated with SARS-CoV-2 infection and its
434 neurological and respiratory manifestations. The SARS-CoV-2 virus enters mainly by air and
435 reaches the lungs through direct ventilation and the CNS through the olfactory and trigeminal
436 nerve, the entry of the virus is facilitated by NRP-1, ACE2 receptors and the protein S
437 activation by TMPRSS2. In the CNS, the virus infects neurons, glial cells, and endothelial
438 cells, increasing the permeability of the BBB, and may cause cerebral edema and intracranial
439 hypertension, as well as neuroinflammation. If the viral infection continues, the damage
440 spreads throughout the body causing heart and systemic failure. This damage is associated
441 with an increase in neuroinflammation, directed by microglia and oligodendrocytes, causing
442 damage to the brain stem, and causing a dysfunctional state of the heart and lung. Likewise,
443 in the lung, due to exacerbated inflammation and intravascular coagulation, respiratory arrest
444 is induced that can lead to the patient death. The inflammation is conducted by the cellular
445 activation through TLR3, 7 and 8 for components from the virus (PAMPS) and subsequent
446 production of pro-inflammatory cytokines (TNF α and IL 1 β) and generation of ROS; those
447 ROS can be able to modify the P2X7 receptor in the brain and activate the inflammasome by
448 the decrease of K⁺. The activation of inflammasome increases the production of IL-6 and
449 pyroptosis.

450 **Figure 2.** Outline of the REVIVAL trial clinical protocol. Initially, patients will be informed
451 about the clinical trial, if they accept and sign the consent, they will be randomized using the
452 Sealed envelope® software. Group A receive DXM intranasally obtaining serum and a swab
453 on days 0, 3, 6 and 10 post treatment. On the other hand, group B receive intravenous DXM,
454 obtaining the same samples on the same days 0,3,6 and 10. Throughout the study, the patients

455 are monitored. Once the results are obtained, these are analyzed to define if exist a
456 statistically difference between groups.

457 **References**

- 458 [1] Medzhitov R. Origin and physiological role of inflammation. *Nature* (2008) 454:428–
459 35.10.1038/nature07201.
- 460 [2] Li YC, Bai WZ, Hashikawa T. The neuroinvasive potential of SARS-CoV2 may play a
461 role in the respiratory failure of COVID-19 patients. *J Med Virol.* 2020;92:552-555
- 462 [3] Zhou Z, Kang H, Li S, Zhao X. Understanding the neurotropic characteristics of SARS-
463 CoV-2: from neurological manifestations of COVID-19 to potential neurotropic
464 mechanisms. *J Neurol.* 2020;267(8):2179-2184
- 465 [4] Ng Kee Kwong, Koy Chong, Puja R. Mehta, Garima Shukla, and Arpan R. Mehta. 2020.
466 “COVID-19, SARS and MERS: A Neurological Perspective.” *Journal of Clinical*
467 *Neuroscience.* Churchill Livingstone. <https://doi.org/10.1016/j.jocn.2020.04.124>.
- 468 [5] Li YC, Bai WZ, Hirano N, Hayashida T, Hashikawa T. Coronavirus infection of rat dorsal
469 root ganglia: ultrastructural characterization of viral replication, transfer, and the early
470 response of satellite cells. *Virus Res.* 2012;163(2):628-635
- 471 [6] Desforges M, Le couponec A, Dubeau P, Bourgouin A, Lajoie L, Dubé M, Talbot PJ.
472 Human coronaviruses and other respiratory viruses: underestimated opportunistic pathogens
473 of the central nervous system? *Viruses* 2019;12(1):14
- 474 [7] Gasmi A, Noor S, Menzel A, Dosa A, Pivina L, Bjoorklund G. Obesity and Insulin
475 Resistance: Associations with Chronic Inflammation, Genetic and Epigenetic Factors. *Curr*
476 *Med Chem* 2020. doi: 10.2174/0929867327666200824112056.

- 477 [8] Liu Danlin, Richardson G, Benli FM, Park C, de Souza JV, Bronowska AK,
478 Spyridopoulos. Inflammation in the cardiovascular system: mechanisms, emerging targets,
479 and novel therapeutic strategies. *Clin Sci (Lond)* 2020; 134(17):2243-2262.
- 480 [9] Meinhardt J, Radke J, Dittmayer C et al. Olfactory transmucosal SARS-CoV-2 invasion
481 as a port of a central nervous system entry in individuals with COVID-19. *Nat Neurosci*
482 2021;24(2):1698-175
- 483 [10] Dos Santos MF, Devalle S, Aran V, Capra D, Roque NR, Coelho-Aguiar JM, Spohr
484 TCLSE, Subilaga JG, Pereira CM, D'Andrea Meira I, Niemeyer Soares Filho P, Moura-Neto
485 V. Neuromechanisms of SARS-CoV-2: A review. *Front Neuroanat.* 2020;14:37
- 486 [11] Somoloni IH, Normandin E, Bhattacharyya S, Mukerji SS, Kellner K, Ali AS, Adams G,
487 Hornick JL, Padera RF Jr, Sabeti P. Neuropathological features of COVID-19. *N Engl J Med*
488 2020;383:989-992
- 489 [12] Paniz-Mondolfi A, Bryce C, Grimes Z, Gordon RE, Reidy J, Lednicky J, Sordillo EM,
490 Fowkes M. Central nervous system involvement by severe acute respiratory syndrome
491 coronavirus-2 (SARS-CoV-2). *J Med Virol.* 2020;92(7):699-702
- 492 [13] Kanberg N, Ashton NJ, Andersson LM, Yilmaz A, Lindh M, Nilsson S, Price RW,
493 Blennow K, Zetterberg H, Gisslén M. Neurochemical evidence of astrocytic and neuronal
494 injury commonly found in COVID-19. *Neurology.* 2020;e1754-e1759.
- 495 [14] Andrews MG, Mukhtar T, Enze UC, Simoneau CR, Perez Y, Mostajo-Radji MA, Wang
496 S, Velmeshev D, Salma J, Kumar GR, Pollen AA, Crouch EE, Ott M, Kriegstein AR. Tropism

497 of SARS-CoV-2 for developing human cortical astrocytes. Version 1. bioRxiv. Preprint.2021
498 Jan 18. Doi:10.1101/2021.01.17.427024

499 [15] Ramani A, Müller L, Ostermann PN, Gabriel E, Abida-Islam P, Müller-Schiffmann A,
500 Mariappan A, Goureau O, Gruell H, Walker A, Andrée M, Hauka S, Houwaart T, Dilther A,
501 Wohlgemuth K, Omran H, Klein F, Wiczorek D, Adams O, Timm J, Korth C, Schaal H,
502 Gopalakrishnan J. SARS-CoV-2 targets neurons of 3D human brain organoids. *EMBO J*.
503 2020;39:e106230

504 [16] Song E, Zhang Ce, Israelow B, Lu-Culligan A, Veites Prado A, Skriabine S, Lu P, Orr-
505 El Weizman L, Lui F, Dai Y, Szigeti-Buck K, Yasumoto Y, Wang G, Castaldi C, Heltke J,
506 Ng E, Wheeler J, Madel Alfajaro M, Levavasseur E, Fontes B, Ravindra NG, van Dijk D,
507 Gunel M, Ring A, Jaffar Kazmi SA, Khang K, Willen CB, Horvath TL, Plu I, Haik S, Thomas
508 JL, Louvi A, Farhadian SF, Huttner A, Seilhean D, Renier N, Bilguvar K, Iwaski A.
509 Neuroinvasion of SARS-CoV-2 in human and mouse brain. *J Exp Med*. 2021;218:e20202135

510 [17] Siddiqi HS, Mehra MR. COVID-19 illness in native and immunosuppressed states: A
511 clinical-therapeutic staging proposal. *J Heart Lung Transplant*. 2020;39(5):405-407

512 [18] Hoffmann, Markus, Hannah Kleine-Weber, Simon Schroeder, Nadine Krüger, Tanja
513 Herrler, Sandra Erichsen, Tobias S. Schiergens, et al. 2020. "SARS-CoV-2 Cell Entry
514 Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor."
515 *Cell* 181 (2): 271-280.e8 <https://doi.org/10.1016/j.cell.2020.02.052>

516 [19] Sungnak W, Huang N, Becavin C, et al. SARS-CoV-2 entry factors are highly expressed
517 in nasal epithelial cells together with innate immune genes. *Nat Med* 2020; 26: 681–687.
518 doi:10.1038/s41591-020-0868-6

519 [20] Cantuti-Castelvetri L, Ojha R, Pedro LD, Djannatian M, Franz J, Kuivanen S, van der
520 Meer F, Kallio K, Kaya T, Anastasina M, Smura T, Levanov L, Szivovicza L, Tobi A, Kallio-
521 Kokko H, Österlund P, Joensuu M, Meunier FA, Butcher SJ, Winkler MS, Mollenhauer B,
522 Helenius A, Gokce O, Teesalu T, Hepojoko J, Vapalahti O, Stadelmann C, Balistreri G,
523 Simons M. Neuropilin-1 facilitates SARS-CoV-2 cell entry and infectivity. *Science*
524 2020;370:856-860.

525 [21] Davies J, Randeve HS, Chatha K, Hall M, Spandidos DA, Karteris E, Kyrou I.
526 Neuropilin-1 as a new potential SARS-CoV-2 infection mediator implicated in the
527 neurologic features and central nervous system involvement of COVID-19. *Mol Med Rep.*
528 2020;22:4221-4226

529 [22] Butowt, R. and Bilinska, K. (2020) SARS-CoV-2: Olfaction, brain infection and the
530 urgent need for clinical samples allowing earlier virus detection. *ACS Chem. Neurosci.* 11
531 (9), 1200– 1203, doi: 10.1021/acchemneuro.0c00172

532 [23] Yap JKY; Moriyama M, Iwasaki A. Inflammasomes and pyroptosis as therapeutic
533 targets for COVID-19. *J Immunol.* 2020;205(2):307-312

534 [24] Freeman TL, Swartz TH. Targeting the NLRP3 inflammasome in severe COVID-19.
535 *Front Immunol.* 2020;11:1518

536 [25] Ribeiro Dem Oliveira-Giacomelli A, Glaser T, Arnaud.Sampaio VF, Andrejew R,
537 Diekmann L, Baranova J, Lameu C, Ratajczak M, Ulrich H. Hyperactivation of P2X7
538 receptors as a culprit of COVID-19 neuropathology. *Mol Psychiatry* 2020;1-16

539 [26] Netland J, Meyerholz DK, Moore S, Cassell M, Perlman S. Severe acute respiratory
540 syndrome coronavirus infection causes neuronal death in the absence of encephalitis in mice
541 transgenic for human ACE2. *J Virol.* 2008;82(15):7264-7275.

542 [27] Wu Y., Xu X., Chen Z., Duan J., Hashimoto K., Yang L. Nervous system involvement
543 after infection with COVID-19 and other coronaviruses. *Brain Behav.*
544 *Immun.* 2020;20 30357-3.

545 [28] Zhou L., Zhang M., Wang J., Gao J. Sars-Cov-2: underestimated damage to nervous
546 system. *Travel Med. Infect. Dis.* 2020 doi: 10.1016/j.tmaid.2020.101642.

547 [29] Sun T., Guan J. Novel coronavirus and central nervous system. *Eur. J. Neurol.* 2020 doi:
548 10.1111/ene.14227.

549 [30] Gandhi S, Srivastava AK, Ray U, Tripathi PP. Is the collapse of the respiratory center
550 in the brain responsible for respiratory breakdown in COVID-19 patients? *ACS Chem*
551 *Neurosci* 2020. <https://doi.org/10.1021/acchemneuro.0c00217>

552 [31] Misra R, Florez-Perdomo WA, Vasquez HE, Moscote-Salazar LR, Agrawal A. SARS-
553 CoV2 and the pathobiology of the respiratory control mechanism in the brainstem. *J Formos*
554 *Med Assoc.* 2020: S0929-6646(20)30347-8

555 [32] Chigr F, Merzouki M, Najimi M. Autonomic brain centers and pathophysiology of
556 COVID-19. *ACS Chem Neurosci* 2020;11(11):1520-1522

557 [33] Cai Y, Hay M, Bishop VS. Synaptic connections and interactions between area postrema
558 and nucleus tractus solitarius. *Brain Res.*1996; 724, 121–124. doi: 10.1016/0006-
559 8993(96)00282-X

560 [34] Goodchild AK, Moon EA. Maps of cardiovascular and respiratory regions of rat ventral
561 medulla: focus on the caudal medulla. *J Chem Neuroanat.* 2009;38(3):209-21

562 [35] Delgado-Roche L, Mesta F. Oxidative stress as key player in severe acute respiratory
563 syndrome coronavirus (SARS-CoV) infection. *Arch Med Res.* 2020; 51:384-387

564 [36] Hamrahian AH, Fleseriu M; AACE adrenal scientific committee. Evaluation and
565 management of adrenal insufficiency in critically ill patients: disease state review. *Endocr*
566 *Pract.* 2017;23(6):716-725

567 [37] Mateos Moreno L, Palacios Garcia N, Estrada Garcia FJ. Adrenal insufficiency in
568 critical patients: new etiopathogenic concepts and therapeutic implications. *Endocrinol*
569 *Diabetes Nutr.* 2017;64(10):557-563.

570 [38] Hamming I, Timens W, Bultuis MLC, Lely AT, Navis GJ, van Goor H. Tissue
571 distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in
572 understanding SARS pathogenesis. *J Pathol.* 2004;203(2):631-637

573 [39] Ding YQ, Wang HJ, Shen H, Li ZG, Geng J, Han HX, Cai JJ, Li X, Kang W, Weng DS,
574 Lu YD, Yao KT. Study on etiology and pathology of severe acute respiratory syndrome.
575 *Zhonghua Bing Li Xue Za Zhi.* 2003;32(3):195-200

576 [40] Leow MKS, Kwek DSK, Ng AWK, Kaw GJL, Lee LSU. Hypocortisolism in survivors
577 of severe acute respiratory syndrome (SARS). *Clin Endocrinol (Oxf).*2005;63:197-202

578 [41] Alzahrani AS, Mukhtar N, Aljomaiah A, Aljamei H, Barkhsh A, Alsudani N, Elsayed
579 T, Alrashidi N, Fadel R, Alqahtani E, Raef H, Butt MI, Sulaiman O. *Endocr Pract.*
580 2021;27(2):83-89

581 [42] Pal R. COVID-19, hypothalamo-pituitary-adrenal axis and clinical implications.
582 *Endocrine.* 2020;68(2):251-252

583 [43] Coutinho AE, Chapman KE. The anti-inflammatory and immunosuppressive effects of
584 glucocorticoids, recent developments and mechanistic insights. *Mol Cell Endocrinol.*
585 2011;335:2-13

586 [44] Van Der Velden VHJ. Glucocorticoids: mechanisms of action and anti-inflammatory
587 potential in asthma. *Mediators Inflamm.* 1998 (7):229-237

588 [45] Busillo JM, Azzam KM, Cidlowski JA. Glucocorticoids sensitize the innate immune
589 system through regulation of the NLRP3 inflammasome. *J Biol Chem.* 2011;286:38703-
590 38713

591 [46] Pinto A, Jacobsen M, Geoghegan PA, Cangelosi A, Cejudo ML, Tironi-Farinati C,
592 Goldstein J. Dexamethasone rescues neurovascular unit integrity from cell damage caused
593 by systemic administration of shiga toxin 2 and lipopolysaccharide in mice motor cortex.
594 *PLoS One* 2013;8:e70020

595 [47] RECOVERY Collaborative Group, Horby P, Lim WS, Emberson JR, Mafham M, Bell
596 JL, Linsell L, et al. Dexamethasone in hospitalized patients with Covid-19- Preliminary
597 report. *N Engl J Med.* 2020 Jul 17.

598 [48] Singh AK, Majumdar S, Singh R, Misra A. Role of corticosteroids in the management
599 of COVID-19: A systemic review and a clinician's perspective. *Diabetes Metab Syndr.*
600 2020;14(5):971-978

601 [49] Selvaraj V, Dappah-Afriyie K, Finn A, Flanigan TP. Short-term dexamethasone in
602 SARS-CoV-2 patients. *R I Med J* (2013). 2020;103(6):39-43

603 [50] Ledford H. Coronavirus breakthrough: dexamethasone is first drug shown to save lives.
604 *Nature.* 2020;582(7813):469

605 [51] Isidori AM, Arnaldi G, Boscaro M, et al. COVID-19 infection and glucocorticoids:
606 update from the Italian Society of Endocrinology Expert Opinion on steroid replacement in
607 adrenal insufficiency. *J Endocrinol Invest.* 2020;43(8):1141-1147. doi:10.1007/s40618-020-
608 01266-w

609 [52] Villar J, Ferrando C, Martínez D, et al. Dexamethasone treatment for the acute res
610 piratory distress syndrome: a multicentre, randomised controlled trial. *Lancet Respir Med.*
611 2020;8(3):267–276. doi:10.1016/S2213-2600(19)30417-5.

612 [53] Russell CD, Millar JE, Baillie JK. Clinical evidence does not support corticosteroid
613 treatment for 2019-nCoV lung injury. *Lancet.* 2020;395(10223):473–475.
614 doi:10.1016/S0140-6736(20)30317-2.

615 [54] Lee N, Allen Chan KC, Hui DS. Effects of early corticosteroid treatment on plasma
616 SARS-associated Coronavirus RNA concentrations in adult patients. *J Clin Virol.*
617 2004;31:304–309.

618 [55] Erdó F, Bors LA, Farkas D, Bajza Á, Gizurarson S. Evaluation of intranasal delivery
619 route of drug administration for brain targeting. *Brain Res Bull.* 2018;143:155-170

620 [56] Meneses G, Cárdenas G, Espinosa A, et al. Sepsis: developing new alternatives to reduce
621 neuroinflammation and attenuate brain injury. *Ann N Y Acad Sci.* 2019;1437(1):43–56.
622 doi:10.1111/nyas.13985

623 [57] Espinosa A, Meneses G, Chavarría A, Mancilla R, Pedraza-Chaverri J, Fleury A,
624 Barcena B, Pérez Osorio IN, Besedovsky H, Arauz A, Fragoso G, Scitutto E. Intranasal
625 dexamethasone reduces mortality and brain damage in a mouse experimental ischemic stroke
626 model. *Neurotherapeutics* 2020 Jul 6. Doi:10.1007/s13311-020-00884-9

627 [58] Meneses G, Gevorkian G, Florentino A, et al. Intranasal delivery of dexamethasone
628 efficiently controls LPS-induced murine neuroinflammation. *Clin Exp Immunol.*
629 2017;190(3):304–314. doi:10.1111/cei.13018

630 [59] Rassy D, Bárcena B, Pérez-Osorio IN, et al. Intranasal Methylprednisolone Effectively
631 Reduces Neuroinflammation in Mice with Experimental Autoimmune Encephalitis. *J*
632 *Neuropathol Exp Neurol.* 2020;79(2):226–237.

633 [60] Zhang Y, Hu S, Wang J, Xue Z, Wang C, Wang N. Dexamethasone inhibits SARS-
634 CoV-2 spike pseudotyped virus viropexis by binding to ACE2. *Virology* 2021;554:83-88

635 [61] Finney LJ, Glaville N, Farne H, et al. Inhaled corticosteroids downregulate the SARS-
636 CoV-2 receptor ACE2 in COPD through suppression of type I interferon. *BioRxiv* 2020.
637 Doi:10.1101/202.06.13.149039

638 [62] Wang T, Zhao Y, Fan F, Hu R, Jin X. Dexamethasone inhibits *S. aureus*-induced
639 neutrophils extracellular pathogen-killing mechanism, possibly through toll-like receptor
640 regulation. *Front Immunol* 2017;8:60. Doi:10.3389/fimmu.2017.00060.

641 [63] Zuo Y, Yalavarthi S, Shi H, Gockman K, Zuo M, Madison JA, et al. Neutrophils
642 extracellular traps (NETs) as markers of disease severity in COVID-19. *medRxiv*.
643 2020:2020.04.09.20059626

644 [64] Middleton EA, He XY, Denorme F, Campbell RA, Ng D, Salvatore SP et al..
645 Neutrophils extracellular traps (NETs) contribute to immunothrombosis in COVID-19 acute
646 respiratory distress syndrome. *Bloods* 2020: blood.2020007008

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